BBA 79306

PHOTOINACTIVATION OF THE THIAMINE TRANSPORT SYSTEM IN SACCHAROMYCES CEREVISIAE WITH 4-AZIDO-2-NITROBENZOYLTHIAMINE

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(Received February 2nd, 1981)

Key words: Photoinactivation; Thiamine transport; 4-Azido-2-nitrobenzoylthiamine; (S. cerevisiae)

A newly synthesized photoreactive thiamine derivative, 4-azido-2-nitrobenzoylthiamine was found to be a competitive inhibitor of the thiamine transport system in $Saccharomyces\ cerevisiae$, exhibiting an apparent K_i of 36 nM. When exposed to visible light, 4-azido-2-nitrobenzoylthiamine irreversibly inactivated the thiamine transport. 4-Azido-2-nitrobenzoylthiamine-dependent photoinactivation of thiamine transport was partially protected by thiamine, but not by the nitrene-trapping reagent p-aminobenzoate. On the other hand, the irradiation of the yeast cells in the presence of 4-azido-2-nitrobenzoylthiamine did not significantly lead to inactivation of the biotin transport system. The results suggest that 4-azido-2-nitrobenzoylthiamine is a specific irreversible inhibitor of the thiamine transport system in $Saccharomyces\ cerevisiae$.

It has been recognized for several years that thiamine enters yeast cells by a carrier-mediated active transport process [1,2,3]. However, little is known concerning the molecular constituents of the membrane which are involved in the transport process. In this paper we wish to report that 4-azido-2-nitrobenzoylthiamine, a newly synthesized derivative of thiamine, inactivates the thiamine transport system in Saccharomyces cerevisiae when exposed to visible light, suggesting that this compound is an effective irreversible inhibitor of the thiamine transport system in yeast cell membrane.

Materials and Methods

Chemicals. [thiazole-2-14C] thiamine hydrochloride (24.3 Ci/mol) and D-[carbonyl-14C] biotin (49 Ci/mol) were obtained from the Radiochemical Centre, U.K. 5-Hydroxyethyl-4-methylthiazole [4] and 4-amino-5-bromomethyl-2-methylpyrimidine hydrochloride. 4-Azido-2-nitrobenzoic acid was synthesized from 4-amino-2-nitrobenzoic acid essentially by the method of Lewis et al. [7], which applies to the

synthesis of 5-azido-2-nitrobenzoic acid from 5-amino-2-nitrobenzoic acid. All other chemicals were purchased from commercial suppliers.

Growth of yeast cells. S. cerevisiae was grown in Wickerham's synthetic medium thiamine was omitted, as described previously [3] After harvesting, yeast cells were washed once with cold water.

Transport assays. The transport of thiamine was determined by the method described previously [2]. The irradiated cell suspensions were centrifuged at $2\,000 \times g$ for 5 min, the cell pellets were washed twice with cold water and finally suspended in 1 ml cold water. The aliquots of the cell suspensions were employed to measure the initial rate of thiamine uptake by the cells; 0.2 ml cell suspension was added to 3.8 ml 50 mM potassium phosphate buffer (pH 5.0) containing 0.1 M glucose and 40 μ l 0.1 mM [thiazole-2-¹⁴C]thiamine which was prewarmed at 37°C. After 2 min at 37°C the radioactivity of 1 ml of the cell suspension was measured as previously reported [2].

Irradiation procedure. Photoinactivation studies were carried out in a 20-ml glass beaker which was kept in ice. The washed yeast cells, suspended in 2 ml 50 mM potassium phosphate buffer (pH 5.0) at the

concentration of 0.5 mg dry weight per ml, were irradiated in the presence of 4-azido-2-nitrobenzoyl-thiamine and other derivatives of thiamine. Photolysis was carried out for 10 min with a Toshiba black light lamp (40 W), 25 cm from the reaction vessel.

4-azido-2-nitrobenzoylthiamine. Synthesis of4-Amino-2-nitrobenzoic acid (5.2 g, 28.5 mmol) was dissolved in 12 M HCl (45 ml); NaNO₂ (3.19 g, 46.2 mmol) dissolved in water (15 ml) was added portionwise with stirring over 60 min at $-5-0^{\circ}$ C; acetic acid (42.7 ml) was added at $-5-0^{\circ}$ C; NaN₃ (3.20 g, 49.2 ms)mmol) dissolved in water (12 ml) was slowly added drop-wise over 1 h at 0-5°C. The reaction mixture was diluted with cold water (56 ml), stirred another 15 min at 0-5°C and further diluted with cold water (200 ml); the resulting precipitate was collected by suction filtration and dried over P2O5 in a desiccator to give pale yellow crystals of 4-azido-2-nitrobenzoic acid weighing in total 5 g (yield 84.2%), m.p. 177-179°C (decomposed), Infrared (KBr) 2120 cm⁻¹ (azido). In the above reaction sequence the addition of NaN₃ and the succeeding procedures were carried out in subdued light.

4-Azido-2-nitrobenzoic acid (0.9 g, 4.32 mmol) was heated with SOCl₂ (3 ml) for 20 min under reflux, concentrated in vacuum at 50°C and traces of SOCl₂ were removed by addition of benzene and alternate evaporation in vacuum, to leave crystalline 4-azido-2-nitrobenzoyl chloride, which was then used without purification. 5-Hydroxyethyl-4-methylthiazole (0.65 g, 4.54 mmol) was added to the chloride, the mixture was refluxed in benzene (2 ml) in the presence of pyridine (0.5 ml) for 1 h, concentrated in vacuum at 50°C and the crystalline residue was recrystallized from ethanol to give 4-methyl-5-[2-(4-azido-2-nitrobenzoyl)ethyl]thiazole (1.25 g, yield) 86.8%), m.p. 94–97°C. Further recrystallization from ethanol gave colorless needles, m.p. 98–99°C.

 $C_{13}H_{11}N_5O_4S$ $(M_r 333.34)$

Calculated: C 46.85, H 3.33, N 21.01 Found: C 46.94; H 3,21, N 20.80

Infrared (KBr) 2130 cm⁻¹ (azido); 1715 cm⁻¹ (carbonyl); 1550, 1375 cm⁻¹ (nitro). Ultraviolet (in ethanol) λ_{max} 250 nm (ϵ = 17800).

A mixture of 4-amino-5-bromomethyl-2-methyl-pyrimidine hydrobromide (1.7 g, 6.01 mmol) and

4-methyl-5-[2-(4-azido-2-nitrobenzoyl)ethyl] thiazole (2.0 g, 6.00 mmol) in *n*-butanol (5 ml) was heated for 75 min under reflux and filtered while hot; the residue was washed with ethanol and then well-washed with acetone to leave a straw-colored powder of 4-azido-2-nitrobenzoylthiamine bromide hydrobromide (0.94 g, crude yield 25.4%), m.p. 204-205°C (decomposed). Recrystallization from ethanol gave colorless crystals, m.p. 207-208°C (decomposed).

 $C_{19}H_{19}BrN_8O_4S \cdot HBr \qquad (M_r 616.30)$

Calculated: C 37.03, H 3.10, N 18.18 Found: C 37.01, H 3.25, N 17.74

Infrared (KBr) 2 130, 2 115 cm⁻¹ (azido); 1 725 cm⁻¹ (carbonyl); 1 550, 1 385 cm⁻¹ (nitro). Ultraviolet (in pH 2 aqueous HCl) λ_{max} 251 nm (ϵ = 29 300).

Results and Discussion

From the transport studies [2] 4-azido-2-nitrobenzoylthiamine was found to be a competitive inhibitor of thiamine uptake by yeast cells and an apparent K_i of 36 nM was calculated from a double reciprocal plot relating initial rate of thiamine uptake to thiamine concentration in the presence of $0.1 \mu M$ 4-azido-2-nitrobenzoylthiamine. Since 4-azido-2nitrobenzoylthiamine contains an aryl azido group in the molecule it was expected that the irradiation of the compound with visible light could affect photolysis of the azido moiety, forming a reactive nitrene which could react covalently with the membrane components functionally involved in the thiamine transport system.

When yeast cells were irradiated with visible light in the presence of 1 μ M 4-azido-2-nitrobenzoylthiamine, their ability to transport thiamine was rapidly lost (Table I). As shown, the initial rate of thiamine transport after irradiation for 10 min decreased to approx. 35% of the original activity. When yeast cells were irradiated in the absence of 4-azido-2-nitrobenzoylthiamine or were kept in the dark with 4-azido-2-nitrobenzoylthiamine for 10 min, their transport activities were almost the same as that of untreated control cells (Table II), indicating that the inactivation by 4-azido-2-nitrobenzoylthiamine is dependent upon irradiation. It should be stressed that the photo-inactivation under these conditions is irreversible

TABLE I

TIME COURSE OF PHOTOINACTIVATION OF THIAMINE TRANSPORT IN *SACCHAROMYCES CEREVISIAE* WITH 4-AZIDO-2-NITROBENZOYLTHIAMINE

Photoinactivation studies were carried out for the indicated time period as described in Materials and Methods.

Minutes of light	Thiamine uptake $(nmol \cdot mg^{-1} \cdot min^{-1})$		
0	6.0		
1	4.3		
2	3.1		
5	2.3		
10	2.1		

since the inactivation could not be reversed by washing the cells free of inactivator. 4-Methyl-5-[2-(4-azido-2-nitrobenzoyl)ethyl] thiazole, an intermediary product with an aryl azido group for the chemical synthesis of 4-azido-2-nitrobenzoylthiamine, was ineffective in inactivating the transport system. This was consistent with previous findings showing that the thiamine transport system in *S. cerevisiae* is specific for the pyrimidine moiety of the thiamine molecule [8,9].

As regards the specificity of 4-azido-2-nitrobenzoylthiamine photoinactivation, the addition of $10 \mu M$ thiamine to inactivation mixture containing $0.5 \mu M$ 4-azido-2-nitrobenzoylthiamine caused at 64.0% increase in thiamine uptake, whereas $10 \mu M$ p-aminobenzoate, a nitrene-trapping reagent [10], produced no protection on 4-azido-2-nitrobenzoylthiamine-dependent photoinactivation of the thiamine transport. These results suggest that nitrene formed from 4-azido-2-nitrobenzoylthiamine bound to the thiamine-specific components (probably thiamine carrier protein) inactivates directly and is not trapped by p-aminobenzoate.

It has been reported that biotin transport in S. cerevisiae occurs by carrier-mediated active transport, which is specifically inactivated with p-nitrophenyl esters of biotin [11]. In contrast to thiamine transport, biotin transport was inactivated only 17.9% by $5\,\mu\mathrm{M}$ 4-azido-2-nitrobenzoylthiamine, whereas thiamine transport was almost completely inactivated.

The data presented here indicate that 4-azido-2-nitrobenzoylthiamine, a structural analog of thiamine which contains a reactive aryl azido group, specifically inactivites the yeast thiamine transport system under the irradiation of visible light. This compound

TABLE II

REQUIREMENT FOR 4-AZIDO-2-NITROBENZOYLTHIA-MINE-DEPENDENT PHOTOINACTIVATION OF THIA-MINE TRANSPORT IN SACCHAROMYCES CEREVISIAE

Photoinactivation studies were carried out as described in Materials and Methods, with or without thiamine derivatives as indicated.

Addition	Concn. (µM)	Irradia- tion	Thiamine uptake (%)
None		_	100
None		+	99.0
4-Azido-2-nitrobenzoyl-			
thiamine	1	_	103.2
	0.5	+	59.9
	1	+	21.7
4-Methyl-5-[2-(4-azido- 2-nitrobenzoyl)ethyl]-			
thiazole	1	+	92.6

may be a potent tool both for identification of the membrane components responsible for active transport of thiamine through the yeast membrane and for elucidating the mechanistic details of the transport process.

Acknowledgement

This work was supported in part by a Grant-in-Aid for Specific Project Research from the Ministry of Education, Science and Culture of Japan.

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